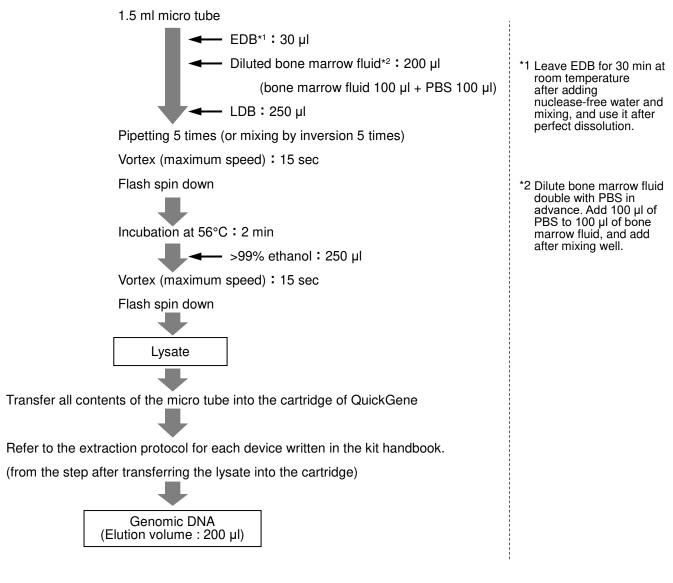
1. Genomic DNA Extraction from Blood of Animal



Genomic DNA Extraction from Bone Marrow Fluid





Results

No Data

Common protocol is usable for the following

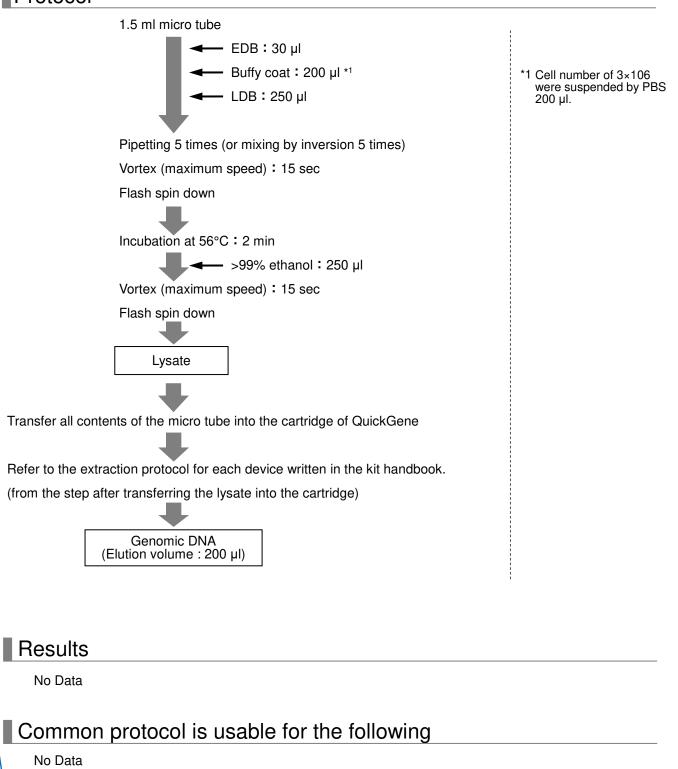
No Data





Genomic DNA Extraction from Buffy Coat

Protocol

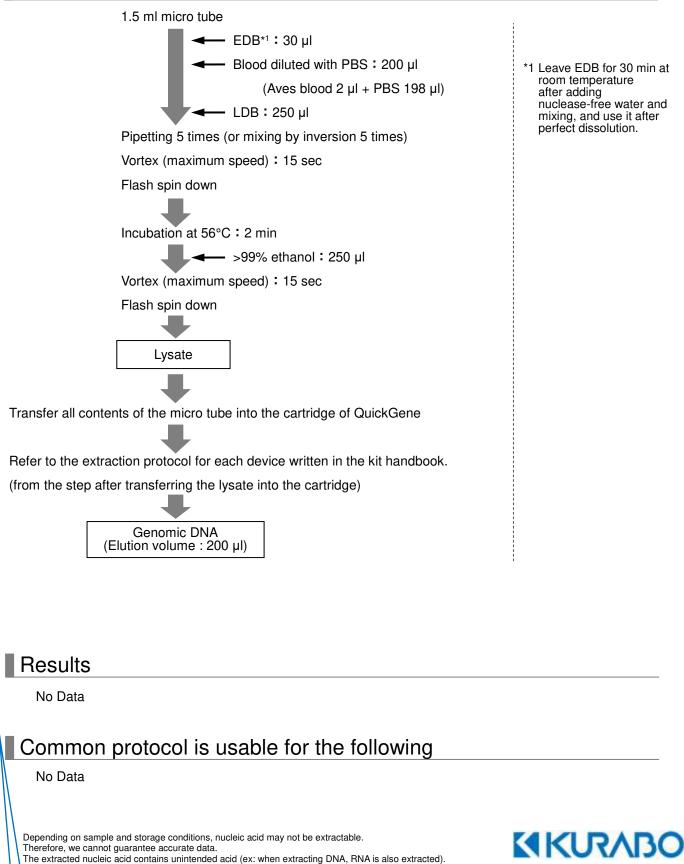






Genomic DNA Extraction from Whole Blood of Aves

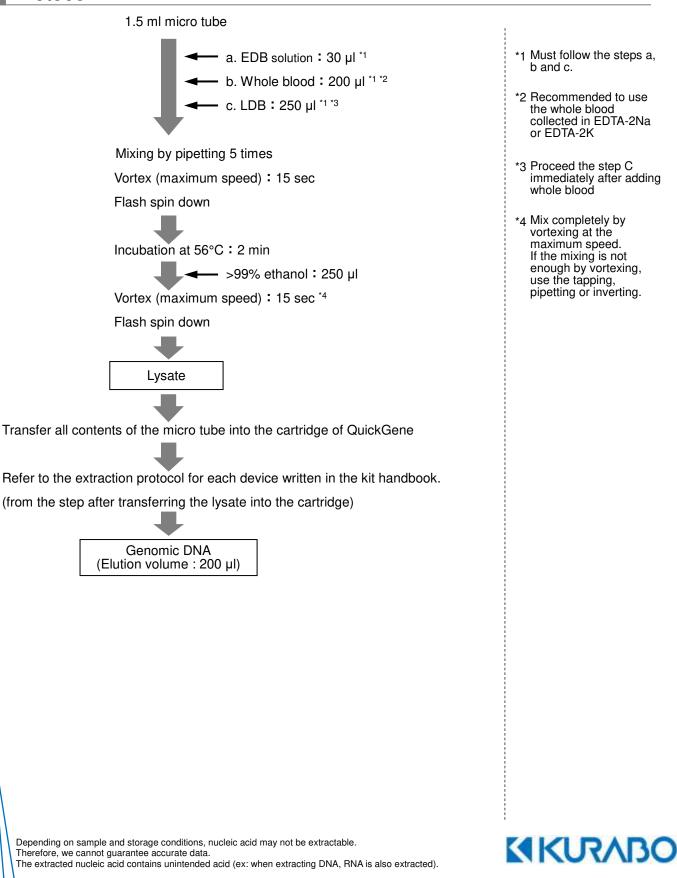
Protocol





Genomic DNA Extraction from Whole Blood of Human

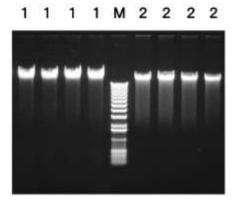
Protocol





Results

Electropherogram



M : 1k bp ladder1 : QuickGene2 : A company (spin method)

The yield of genomic DNA (Sample: 200µl of human whole blood)

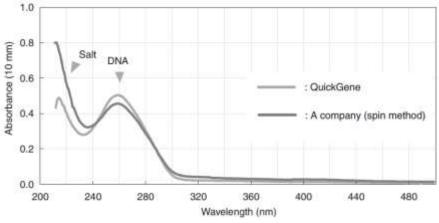
(µg)	Average	Sample 1	Sample 2	Sample 3	Sample 4	Sample 5
QuickGene	5.9	7.2	5.3	5.9	5.5	5.5
A company (spin method)	4.5	6.3	4.4	5.2	3.2	3.6

Protein contamination: A260/280

	Average	Sample 1	Sample 2	Sample 3	Sample 4	Sample 5
QuickGene	1.94	1.91	1.94	1.96	1.91	1.96
A company (spin method)	1.84	1.86	1.82	1.80	1.87	1.86

Chaotropic salt contamination: A260/230

	Average	Sample 1	Sample 2	Sample 3	Sample 4	Sample 5
QuickGene	1.61	1.76	1.69	1.43	1.76	1.42
A company (spin method)	1.12	1.21	0.89	1.07	1.24	1.21



Hemoglobin contamination : A400

	Average	Sample 1	Sample 2	Sample 3	Sample 4	Sample 5
QuickGene	0.036	0.023	0.032	0.070	0.031	0.025
A company (spin method)	0.054	0.076	0.040	0.085	0.026	0.043

Depending on sample and storage conditions, nucleic acid may not be extractable.

Therefore, we cannot guarantee accurate data.

The extracted nucleic acid contains unintended acid (ex: when extracting DNA, RNA is also extracted).

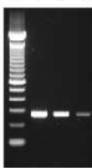




Other

PCR

M 1 2 3

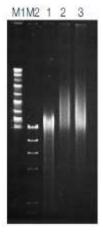


Serial dilution of isolated genomic DNA was used for PCR template to amplify p53 exon6 gene.

PCR amplification was performed successfully by using 0.1ng/µl genomic DNA.

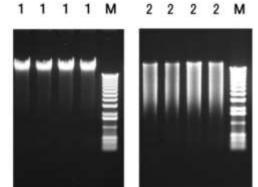
- M: 100 bp ladder
- 1 : Genomic DNA 10ng/µl
- 2 : Genomic DNA 1ng/µl
- 3 : Genomic DNA 0.1ng/µl

Pulsed-field electrophoresis



- The use of QuickGene-810 (automatic nucleic-acid isolation system) and QuickGene DNA whole blood kit S enables the isolation of long genomic DNA same as manual method using phenol / chloroform.
- M1 : MidRange PFG Marker II
- M2 : *Hin*d III digest
- 1 : Comparison method using spin column (<~70kb)
- 2 : Using QuickGene isolation system and reagents (<~140kb)
- 3 : Manual method using phenol / chloroform (<~140kb)

Restriction Enzyme Digestion



The eluted genomic DNA sample had been digested with *Eco*R I.

The success of enzyme digestion is shown by the comparison of lane1 and 2.

- M : 1k bp ladder 1 : Before digestion
- 2 : After digestion using EcoR I





•

Next Generation Sequencing (Exsome sequence analysis)

Genome DNA extracted from whole blood by QuickGene was evaluated for Next Generation Sequencing Analysis, and confirmed it is suitable for NGS.

1	Non-redundant reads (de-duplicated by Picard tools)	Number of non-redundant reads	132,541,212
2	Non-redundant unique reads (uniquely mapped to human genome)	Number of unique reads mapped in human genome	116,879,297
3 (2÷1)	% Non-redundant unique reads (out of non-redundant reads)	Ratio of non-redundant unique reads to non-redundant reads (% for non-redundant reads)	88.2%
4	Target regions (bp)	Number of bases in target regions	62,085,286
5	Number of target genotypes (more than 10X)	Bases covered more than 10x coverage	56,460,863
6 (5÷4)	% Coverage of target region (more than 10X)	Percent bases covered more than 10x coverage	90.9%
7	Mean depth of target regions (X)	Average coverage of target regions	115.8

The quality of DNA library from QuickGene sample was sufficient for NGS with minimal sequence bios and was reliable enough with high depth sequencing across target region.

Common protocol is usable for the following

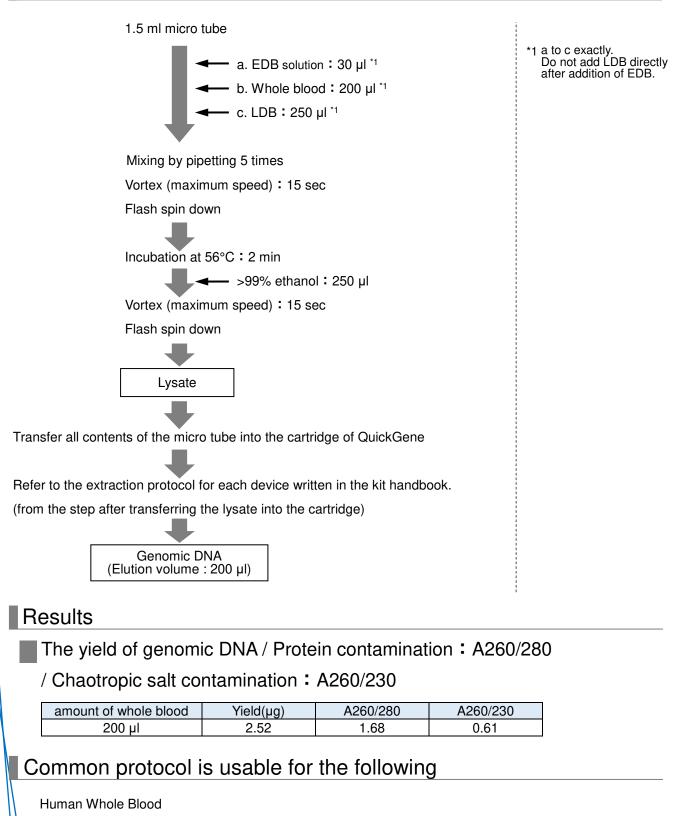
Canine Whole Blood





Genomic DNA Extraction from Whole Blood of Canine

Protocol







DA-a-6

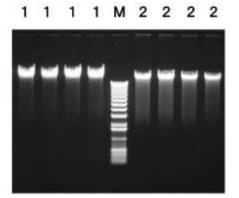
Large-scale Genomic DNA Extraction from Whole Blood of Human

Protocol 15 ml tube *1 Must follow the steps a, a. EDB solution : 300 µl *1 b and c. b. Whole blood : 2 ml *1 *2 *2 Recommended to use - c. LDB : 2.5 ml *1 *3 the whole blood collected in EDTA-2Na or EDTA-2K Mix the sample with shaking upside-down intensely 10 times *3 Proceed the step C Vortex (maximum speed) : 15 sec immediately after adding whole blood *4 Mix completely by Incubation at 56°C: 5 min vortexing at the maximum speed. → >99% ethanol : 2.5 ml If the mixing is not enough by vortexing, Mix the sample with shaking upside-down intensely 10 times use the tapping, pipetting or inverting. Vortex (maximum speed) : 15 sec *4 Lysate Transfer all contents of the micro tube into the cartridge of QuickGene Refer to the extraction protocol for each device written in the kit handbook. (from the step after transferring the lysate into the cartridge) Genomic DNA (Elution volume : 500 µl) Depending on sample and storage conditions, nucleic acid may not be extractable. KKURABO Therefore, we cannot guarantee accurate data. The extracted nucleic acid contains unintended acid (ex: when extracting DNA, RNA is also extracted).



Results

Electropherogram



M : 1k bp ladder 1 : QuickGene 2 : A company (spin method)

The yield of genomic DNA (Sample: 200µl of human whole blood)

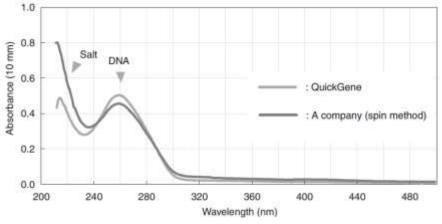
(µg)	Average	Sample 1	Sample 2	Sample 3	Sample 4	Sample 5
QuickGene	5.9	7.2	5.3	5.9	5.5	5.5
A company (spin method)	4.5	6.3	4.4	5.2	3.2	3.6

Protein contamination: A260/280

	Average	Sample 1	Sample 2	Sample 3	Sample 4	Sample 5
QuickGene	1.94	1.91	1.94	1.96	1.91	1.96
A company (spin method)	1.84	1.86	1.82	1.80	1.87	1.86

Chaotropic salt contamination: A260/230

	Average	Sample 1	Sample 2	Sample 3	Sample 4	Sample 5
QuickGene	1.61	1.76	1.69	1.43	1.76	1.42
A company (spin method)	1.12	1.21	0.89	1.07	1.24	1.21



Hemoglobin contamination : A400

	Average	Sample 1	Sample 2	Sample 3	Sample 4	Sample 5
QuickGene	0.036	0.023	0.032	0.070	0.031	0.025
A company (spin method)	0.054	0.076	0.040	0.085	0.026	0.043

Depending on sample and storage conditions, nucleic acid may not be extractable.

Therefore, we cannot guarantee accurate data.

The extracted nucleic acid contains unintended acid (ex: when extracting DNA, RNA is also extracted).

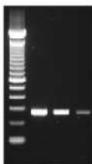




Other

PCR

M 1 2 3



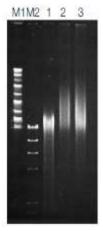
•

Serial dilution of isolated genomic DNA was used for PCR template to amplify p53 exon6 gene.

PCR amplification was performed successfully by using 0.1ng/µl genomic DNA.

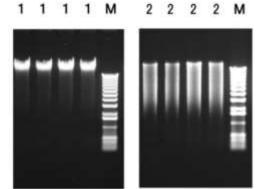
- M: 100 bp ladder
- 1 : Genomic DNA 10ng/µl
- 2 : Genomic DNA 1ng/µl
- 3 : Genomic DNA 0.1ng/µl

Pulsed-field electrophoresis



- The use of QuickGene-810 (automatic nucleic-acid isolation system) and QuickGene DNA whole blood kit S enables the isolation of long genomic DNA same as manual method using phenol / chloroform.
- M1 : MidRange PFG Marker II
- M2 : *Hin*d III digest
- 1 : Comparison method using spin column (<~70kb)
- 2 : Using QuickGene isolation system and reagents (<~140kb)
- 3 : Manual method using phenol / chloroform (<~140kb)

Restriction Enzyme Digestion



The eluted genomic DNA sample had been digested with $\ensuremath{\textit{Eco}}\xspace{\mathsf{R}}$ I.

The success of enzyme digestion is shown by the comparison of lane1 and 2.

- M : 1k bp ladder 1 : Before digestion
- 2 : After digestion using EcoR I



uickGene

•

Next Generation Sequencing (Exsome sequence analysis)

Genome DNA extracted from whole blood by QuickGene was evaluated for Next Generation Sequencing Analysis, and confirmed it is suitable for NGS.

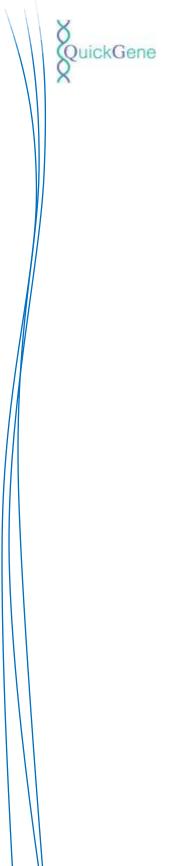
1	Non-redundant reads (de-duplicated by Picard tools)	Number of non-redundant reads	132,541,212
2	Non-redundant unique reads (uniquely mapped to human genome)	Number of unique reads mapped in human genome	116,879,297
3 (2÷1)	% Non-redundant unique reads (out of non-redundant reads)	Ratio of non-redundant unique reads to non-redundant reads (% for non-redundant reads)	88.2%
4	Target regions (bp)	Number of bases in target regions	62,085,286
5	Number of target genotypes (more than 10X)	Bases covered more than 10x coverage	56,460,863
6 (5÷4)	% Coverage of target region (more than 10X)	Percent bases covered more than 10x coverage	90.9%
7	Mean depth of target regions (X)	Average coverage of target regions	115.8

The quality of DNA library from QuickGene sample was sufficient for NGS with minimal sequence bios and was reliable enough with high depth sequencing across target region.

Common protocol is usable for the following

Canine Whole Blood





QuickGene DNA whole blood kit L (DB-L)

